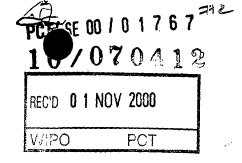
# PRV 5000/ A67

PATENT- OCH REGISTRERINGSVERKET Patentavdelningen





## Intyg Certificate

Härmed intygas att bifogade kopior överensstämmer med de handlingar som ursprungligen ingivits till Patent- och registreringsverket i nedannämnda ansökan.

This is to certify that the annexed is a true copy of the documents as originally filed with the Patent- and Registration Office in connection with the following patent application.

- (71) Sökande AstaCarotene AB, Gustavsberg SE Applicant (s)
- (21) Patentansökningsnummer 9903336-7 Patent application number
- (86) Ingivningsdatum
  Date of filing

1999-09-17

Stockholm, 2000-10-20

För Patent- och registreringsverket For the Patent- and Registration Office

Um ta Todervall

Avgift Fee

PRIORITY
DOCUMENT
SUBMITTED OR TRANSMITTED IN
COMPLIANCE WITH RULE 17.1(a) OR (b)

29295/BN

5

10

15

20

25

30

#### DNA construct and its use.

The present invention relates to a new DNA construct for transformation into oilseed plants. The DNA construct comprises nucleotide sequences encoding peptides with enzyme activities necessary for the high-level production and esterification of keto group-containing xanthophylls in oilseed plants.

#### Background of the invention

Carotenoids are produced *de novo* by plants, fungi, algae and some bacteria. A number of biosynthetic steps are needed for the biological production of the carotenoids. There are two chemically different groups of carotenoids, namely carotenes containing only carbon and hydrogen molecules and xanthophylls containing oxygen in the molecule in addition to carbon and hydrogen.

The xanthophylls, and particularly astaxanthin (3,3'-dihydroxy-β-β-carotene-4,4'-dione), are often colored pigments and are used as such or as anti-oxidants.

Carotenes are biological precursors for the production of the oxygen-containing xanthophylls. There are two types of enzymes responsible for the introduction of hydroxy groups and keto groups into the carotenes, namely hydroxylases and ketolases, respectively.

The keto group-containing xanthophyll astaxanthin, which has keto and hydroxy groups, is biosynthetically produced from beta-carotene.

Large-scale production of xanthophylles from natural sources is at present performed by AstaCarotene AB, Gustavsberg, Sweden, by cultivation of the alga *Haematococcus* pluvialis for the production of astaxanthin in esterified form.

It would be desirable to be able to produce keto group-containing xanthophylls particularly astaxanthin, in oilseed plants. Oilseed plants have naturally  $\beta$ -carotene hydroxylases but lack  $\beta$ -carotene C-4-oxygenase enzymes or ketolases.

#### Description of the invention

The present invention provides DNA constructs enabling and promoting production of keto group containing xanthophylls, especially astaxanthin, in oilseed plants, such as rape, sunflower, soybean and mustard. The DNA construct is transformed into the oilseed plant cell for expression of a protein or fused protein which has an enzyme activity enabling keto group insertion into a carotene or hydroxy carotene for the biosynthetic production of a keto group containing xanthophyll, such as cantaxanthin ( $\beta$ , $\beta$ -carotene-4,4'-dione) and/or astaxanthin. Use is thus made of the biosynthetic pathway of the oilseed plant to



produce carotenoids. The naturally occurring synthesis of carotenoids involves a number of enzymes, namely 1-D-deoxyxylulose 5-phosphate synthase, isopentenyl pyrophosphate:dimethylallyl pyrophosphate isomerase, geranylgeranyl pyrophosphate synthase, phytoene synthase, phytoene desaturase, zeta-carotene desaturase, lycopene beta-cyclase, β-carotene hydroxylase, and β-carotene C-4-oxygenase. Genes coding for peptides having these enzymatic activities may be inserted into the DNA construct of the invention, one or several per construct, to promote high-level production in the transgenic oilseed plant. In case only one enzyme coding gene is inserted per plant, two or more plants may be sexually interbred to produce plants containing all the desired enzyme activities.

5

10

15

20

25

30

Thus, the present invention is directed to a DNA construct comprising in the 5' to 3' direction of transcription operably linked a promoter region directing transcription to the seed of an oilseed plant, a nucleotide sequence coding for at least one peptide with enzyme activity necessary for keto group containing xanthophyll production and esterification in an oilseed plant and a transcriptional termination region.

In a preferred embodiment of the invention the DNA construct additionally comprises between the promoter region and the nucleotide sequence coding for at least one peptide with enzyme activity a nucleotide sequence coding for a transit peptide directing the translated fusion polypeptide to the chloroplast of the oilseed plant.

The DNA construct is preferably such that the promoter is a napin promoter, the peptide with enzyme activity necessary for keto group containing xanthophyll production is selected from the group consisting of peptides with 1-D-deòxyxylulose 5-phosphate synthase, isopentenyl pyrophosphate:dimethylallyl pyrophosphate isomerase, geranylgeranyl pyrophosphate synthase, phytoene synthase, phytoene desaturase, zeta-carotene desaturase, lycopene beta-cyclase,  $\beta$ -carotene hydroxylase, and  $\beta$ -carotene C-4-oxygenase activity. To promote esterification of astaxanthin a nucleotide sequence coding for a peptide with acyl transferase activity may be included in the group.

In a preferred embodiment of the DNA construct according to the invention the nucleotide sequence coding for a peptide with enzyme activity is a nucleotide sequence coding for a N-terminally truncated  $\beta$ -carotene C-4-oxygenase gene from the alga Haematococcus pluvialis.

An example of the DNA construct of the invention is presented in the sequence listing as SEQ ID NO:1 and in Fig.1.



The present invention is also directed to a transgenic oilseed plant cell comprising the DNA construct of the invention, and preferably the oilseed plant is selected from the group consisting of rape, sunflower, soybean and mustard.

The invention is additionally directed to transgenic oilseed plant-produced xanthophyll, e.g. canthaxanthin and astaxanthin.

A preferred aspect of the invention is directed to transgenic oilseed plantproduced astaxanthin esters.

The present invention will now be illustrated with reference to the DNA construct disclosed in the sequence listing and in Fig.1, and the following description of embodiments. However, the invention is not limited to these exemplifications.

### Short description of the drawings

10

15

20

25

30

Fig.1 illustrates the nucleotide sequence of the DNA construct comprising the napin promoter, the chloroplast localization signal, the N-terminally truncated  $\beta$ -carotene C-4-oxygenase gene and the termination sequence, and the deduced amino acid sequences of the transit peptide and the  $\beta$ -carotene C-4-oxygenase.

# **Description of embodiments**

The invention is illustrated by production of astaxanthin in the seed of oilseed rape. The astaxanthin produced in the seed of the transgenic plant is extracted as part of the extracted oil. By use of conventionally used protocols for Agrobacterium tumefaciens mediated transformation such as described by (Hoekema et al. 1983, An et al. 1986, Fry et al. 1987, DeBlock et al. 1988, Radke et al. 1988, or Moloney et al. 1989) transgenic plants are produced having a chimeric DNA construct that is genetically inherited and is able to produce astaxanthin. The nucleotide sequence of the chimeric DNA construct consist of four parts of different genetic origin namely: (1) a promoter, (2) a localization signal, (3) a β-carotene C-4-oxygenase coding region and (4) a termination sequence.

The napin promoter directs transcription to the seed of oilseed rape (Stålberg et al 1996). This promoter was coupled to a localization signal similar but not identical to a transit peptide (TP) of Rbcs1a (Krebbers, 1988) that directs the translated product of a fused gene to the chloroplast. The promoter and the TP sequence were ligated to a part of the coding sequence of a ketolase gene BCK (Kajiwara et al. 1995). This enzyme oxygenates β-carotene to canthaxanthin, (Fraser et al. 1997). The chimeric DNA construct was then coupled to a suitable termination sequence, e.g. that of the Agrobacterium tumefaciens nopaline synthase gene (the nos 3' end)(Bevan et al. 1983), as illustrated in Fig.1.

#### Cellular storage of Astaxantin

5

10

15

25

The storage of large amounts of free astaxanthin in plants will be difficult due to toxic effects of the molecule as it intercalates in the plant membranes. An effective esterification of astaxanthin to fatty acids enables storage of the esterified molecules in triacylglycerol containing oleosomes. Thus, an acyl transferase can be claimed to be of fundamental importance for the process, as is proteins that can mediate transport of different forms of astaxanthin from the chloroplast to the vesicles.

Sequences and oligonucleotides used in the construction of the DNA construct

1. Napin promoter (GeneBank ACCESSION No. J02798)

This promoter sequence, a 1145 base pair fragment including the 5' leader sequence has a unique HindIII site at the 5' end. The 3' end was synthesized with an additionally 6 nucleotide BamHI site.

2. Transit peptide similar to RBCS1a (GeneBank ACCESSION No. X13611, X14565)

The transit peptide (TP) was amplified by PCR from -28 to the end of the transit cleavage aa=54/55 site of the Rbcs1a gene. The 5' end was synthesized with a BamHI site and similarly the 3' sequence was synthesized with a XbaI site. The two following oligonucleotides were used for the PCR amplification.

#### BamHI

20 5' primer: TP1 5'AGAC GGATCC TCAGTCACACAAAGAGTA 3'

SacI XbaI

3' primer: TP2 5'GTTC GAGCTC TCTAGA CATGCAGTTAACGC 3'

3. BCK (\(\beta\)-carotene C-4 oxygenase) (Genebank ACCESSION No. D45881)

The BCK fragment was amplified by PCR including a 5' XbaI site and was ligated to the TP already described. The 5' primer (BCK1) used for PCR, is homologous to the BCK sequence from nucleotide 264 and the 3' oligonucleotide (Ax40) ends with a stop codon and was synthesized with a SacI restriction site for cloning. The synthesized fragment was fused to the TP as shown in Fig 1.

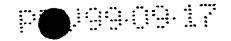
30 Oligonucleotides used for PCR:

XbaI

5' primer: BCK1 5'ACAG TCTAGA ATGCCATCCGAGTCGTCA 3'

SacI

3'primer: AX40 5'CACCGAGCTCCATGACACTCTTGTGCAGA 3'



# Description f SEQ ID NO:1 and SEQ ID NO:2

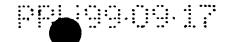
The sequences shown i Fig.1 are the same as the two sequences which are shown in the sequence listing.

The SEQ ID NO:1 is a nucleotide sequence composed of the following features:

5	Nucleotide No.							
	Cloning site HindIII	1-6						
	Napin Promoter	1–1145						
	Cloning site BamHI	1146-1151						
	Transit peptide leader	1152-1178						
10	Transit peptide coding	1179-1347						
	Cloning site Xbal	1348-1353						
	β-carotene C-4-oxygenase	1354-2217						
	$\beta$ -carotene C-4-oxygense 3' untranslated	2218-2266						
	Cloning site SacI	2267-2272						
15	Nopaline synthetase termination	2273-2536						
	Cloning site EcoRI	2538-2543						

The SEQ ID NO: 2 is a deduced amino acid sequence of the fusion protein of the transit peptide and the peptide with  $\beta$ -carotene C-4-oxygenase activity.

::::



#### References

30

An G, Watson BD, Chiang CC (1986), Transformation of tobacco, tomato, potato and Arabidopsis-thaliana using a binary vector system. Plant Physiology 81 (1) 301-305.

Bevan M, Barnes WM and Chilton MD (1983). Structure and transcription of the nopaline synthase gene region of T-DNA. Nucleic Acids Res. 11 (2), 369-385.

DeBlock M, DeBrouwer D, Tenning P (1989). Transformation of Brassica napus and Brassica oleracea using Agrobacterium turnefaciens and the expression of the BAR and NEO genes in transgenic plants Plant Physiology 91:2, 694-701.

Fraser PD, Miura Y, Misawa N, (1997). In vitro characterization of astaxanthin biosynthetic enzymes. J Biol Chem. Mar 7;272(10):6128-35.

Fry J, Barnason A, and Horsch RB, (1987). Transformation of Brassica napus with Agrobacterium tumefaciens based vectors. Plant Cell Reports 6:321-325.

Hoekema A, Hirsch PR, Hooykas PJJ Schilperoort, (1983). A binary vector strategy based on separation of vir and T-region of the Agrobacterium tumefaciens Ti-plasmid. Nature vol 303, 179-180.

Josefsson LG, Lenman M, Ericson ML and Rask L, (1987). Structure of a gene encoding the 1.7 S storage protein, napin, from Brassica napus. J. Biol. Chem. 262 (25), 12196-12201.

Kajiwara S, KakizonoT, Saito T, Kondo K, OhtaniT, Nishio N, Nagai S and Misawa N. (1995). Isolation and functional identification of a novel cDNA for astaxanthin biosynthesis from Haematococcus pluvialis, and astaxanthin synthesis in Escherichia coli Plant Mol. Biol. 29 (2), 343-352.

Krebbers E, Seurinck J, Herdies L, Cashmore AR and Timko MP, (1988). Four genes in two diverged subfamilies encode the rubulose-1, 5-bisphosphate carboxylase small subunit polypeptides of Arabidopsis thaliana Plant Mol. Biol. 11, 745-759.

Moloney M, Walker JM and Sharma KK, (1989). High efficiency transformation of Brassica napus using Agrobacterium vectors. Plant Cell Reports 8:238-242.

Radke SE, Andrews BM, Moloney MM, Crouch ML, Kridl JC, Knauf VC (1988), Transformation of Brassica napus using Agrobacterium tumefaciens – Developmentally regulated Expression of a reintroduced napin gene. TAG, 75: (5) 685-694.

Pua E-C, Mehra-Palta A, Nagy F and Chua N-H, (1987). Transgenic plants of Brassica napus. Biotechnology vol 5, 815-817.

Stålberg K, Ellerstöm M, Ezcurra I, Ablov S, Rask L (1996). Disruption of an overlapping E-box/ABRE motif abolished high transcription of the napA storage-protein promoter in transgenic Brassica napus seeds. Planta 199(4):515-9.

#### SEQUENCE LISTING

```
<110> AstaCarotene AB
<120> DNA construct and its use
<130> 29295-AstaCarotene
 <140>
<141>
<160> 2
<170> PatentIn Ver. 2.1
<210> 1
<211> 2543
<212> DNA
<213> Artificial Sequence
<220>
<223> Description of Artificial Sequence: napin promoter
      + choroplast localization signal + beta-carotene C-4 oxygenase
      coding sequence + termination sequence
<220>
<221> promoter
<222> (1)..(1145)
<220>
<221> transit_peptide
<222> (1179)..(1347)
<220>
<221> CDS
<222> (1179)..(2217)
<220>
<221> terminator
<222> (2273)..(2536)
<400> 1
aagetttett categgtgat tgatteettt aaagaettat gtttettate ttgettetga 60
ggcaagtatt cagttaccag ttaccactta tattctggac tttctgactg catcctcatt 120
tttccaacat tttaaatttc actattggct gaatgcttct tctttgagga agaaacaatt 180
cagatggcag aaatgtatca accaatgcat atatacaaat gtacctcttg ttctcaaaac 240
atctatcgga tggttccatt tgctttgtca tccaattagt gactacttta tattattcac 300
tectetttat tactatttte atgegaggtt gecatgtaca ttatatttgt aaggattgac 360
gctattgagc gtttttcttc aattttcttt attttagaca tgggtatgaa atgtgtgtta 420
gagttgggtt gaatgagata tacgttcaag tgaagtggca taccgttctc gagtaaggat 480
gacctaccca ttcttgagac aaatgttaca ttttagtatc agagtaaaat gtgtacctat 540
```

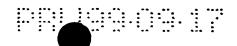
aactcaaatt cgattgacat gtatccattc aacataaaat taaaccagcc tgcacctgca 600 tecacattre aagtattre aaaccetteg geteetatee acceggeteta acaagacega 660 ttccgaattt ggaagatttt gactcaaatt cccaatttat attgaccgtg actaaatcaa 720 ctttaacttc tataattctg attaagctcc caatttatat tcccaacggc actacctcca 780 aaatttatag actctcatcc ccttttaaac caacttagta aacgtttttt tttttaattt 840 tatgaagtta agtttttacc ttgtttttaa aaagaatcgt tcataagatg ccatgccaga 900 acattageta caegttacae atageatgea geogeggaga attgttttte ttegecaett 960 gteactecet teaaacacet aagagettet eteteacage acacacatae aateacatge 1020 gtgcatgcat tattacacgt gategecatg caaateteet ttatageeta taaattaact 1080 catccgcttc actctttact caaaccaaaa ctcatcaata caaacaagat taaaaacata 1140 cacgaggate etcagteaca caaagagtaa agaagaaca atg get tee tet atg 1194 Met Ala Ser Ser Met ctc tot toe get act atg gtt gee tot eeg get eag gee act atg gte 1242 Leu Ser Ser Ala Thr Met Val Ala Ser Pro Ala Gln Ala Thr Met Val 20 10 15 get cet the aac gga ett aag tee tee get gee the cea gee ace ege 1290 Ala Pro Phe Asn Gly Leu Lys Ser Ser Ala Ala Phe Pro Ala Thr Arg 25 30 1338 aag get aac aac gac att act tee ate aca age aac gge gga ege gtt Lys Ala Asn Asn Asp Ile Thr Ser Ile Thr Ser Asn Gly Gly Arg Val 1386 aac tgc atg tct aga atg cca tcc gag tcg tca gac gca gct cgt cct Asn Cys Met Ser Arg Met Pro Ser Glu Ser Ser Asp Ala Ala Arg Pro 55 60 geg cta aag cac gee tac aaa eet eea gea tet gae gee aag gge ate 1434 Ala Leu Lys His Ala Tyr Lys Pro Pro Ala Ser Asp Ala Lys Gly Ile 70 75 1482 acg atg gcg ctg acc atc att ggc acc tgg acc gca gtg ttt tta cac Thr Met Ala Leu Thr Ile Ile Gly Thr Trp Thr Ala Val Phe Leu His 1530 gea ata ttt caa atc agg cta ccg aca tcc atg gac cag ctt cac tgg Ala Ile Phe Gln Ile Arg Leu Pro Thr Ser Met Asp Gln Leu His Trp 105 110 ttg cct gtg tcc gaa gcc aca gcc cag ctt ttg ggc gga agc agc agc 1578 Leu Pro Val Ser Glu Ala Thr Ala Gln Leu Leu Gly Gly Ser Ser Ser 120 125 130 1626 cta ctg cac atc gct gca gtc ttc att gta ctt gag ttc etg tac act Leu Leu His Ile Ala Ala Val Phe Ile Val Leu Glu Phe Leu Tyr Thr

135

140

145

.



ggt Gly 150	cta Leu	ttc Phe	atc Ile	acc Thr	aca Thr 155	cat His	<b>Asp</b>	gca Ala	atg Met	cat His 160	ggc Gly	acc Thr	ata Ile	gct Ala	ttg Leu 165	1674
															ctg Leu	1722
							atg Met								cac His	1770
							999 Gly 205								gga Gly	1818
							ttc Phe									1866
							ctg Leu									1914
_	-				_	_	aat Asn			_		_	_	_	_	1962
		_		_		_	ctc Leu							_		2010
							gca Ala 285									2058
Phe	Arg 295	Ala	Lys	Thr	Ser	Glu 300	gca Ala	Ser	Asp.	Val	Met 305	Ser	Phe	Leu	Thr	2106
Сув 310	Tyr	His	Phe	Asp	<b>Leu</b> 315	His	tgg Trp	Glu	His	His 320	Arg	Trp	Pro	Phe	Ala 325	2154
Pro													Arg			2202
gtg Val		Ala			tgac	ctgg	tc c	ctcc	gctg	g tg	accc	agcg	tct	gcac	aag	2257
agtg	tcat	gg a	gete	gaat	t tc	cccg	atcg	ttc	aaac	att	tggc	aata	aa g	tttc	ttaag	2317
															gttaa	
															attag	
agtocogcaa ttatacattt aatacgogat agaaaacaaa atatagogog caaactagga taaattatog ogogoggtgt catctatgtt actagatogg gaatto							2543									
		-			_	-										_

<210> 2

<211> 346

<212> PRT

<213> Artificial Sequence

<223> Description of Artificial Sequence: deduced fusion protein of transit peptide + peptide with beta-carotene C-4 oxygenase activity

<400> 2

Met Ala Ser Ser Met Leu Ser Ser Ala Thr Met Val Ala Ser Pro Ala

1 5 10 1!

Gln Ala Thr Met Val Ala Pro Phe Asn Gly Leu Lys Ser Ser Ala Ala 20 25 30

Phe Pro Ala Thr Arg Lys Ala Asn Asn Asp Ile Thr Ser Ile Thr Ser
35 40 45

Asn Gly Gly Arg Val Asn Cys Met Ser Arg Met Pro Ser Glu Ser Ser 50 55 60

Asp Ala Arg Pro Ala Leu Lys His Ala Tyr Lys Pro Pro Ala Ser 65 70 75 80

Asp Ala Lys Gly Ile Thr Met Ala Leu Thr Ile Ile Gly Thr Trp Thr 85 90 95

Ala Val Phe Leu His Ala Ile Phe Gln Ile Arg Leu Pro Thr Ser Met
100 105 110

Asp Gln Leu His Trp Leu Pro Val Ser Glu Ala Thr Ala Gln Leu Leu 115 120 125

Gly Ser Ser Ser Leu Leu His Ile Ala Ala Val Phe Ile Val Leu 130 135 140

Glu Phe Leu Tyr Thr Gly Leu Phe Ile Thr Thr His Asp Ala Met His 145 150 155 160

Gly Thr Ile Ala Leu Arg His Arg Gln Leu Asn Asp Leu Leu Gly Asn 165 170 175

Ile Cys Ile Ser Leu Tyr Ala Trp Phe Asp Tyr Ser Met Leu His Arg 180 185 190

Lys His Trp Glu His His Asn His Thr Gly Glu Val Gly Lys Asp Pro 195 200 205

Asp Phe His Lys Gly Asn Pro Gly Leu Val Pro Trp Phe Ala Ser Phe 210 215 220

Met Ser Ser Tyr Met Ser Leu Trp Gln Phe Ala Arg Leu Ala Trp Trp 225 230 235 240

Ala Val Val Met Gln Met Leu Gly Ala Pro Met Ala Asn Leu Leu Val 245 250 255 Phe Met Ala Ala Ala Pro Ile L u Ser Ala Phe Arg Leu Phe Tyr Phe 260 265 270

Gly Thr Tyr Leu Pro His Lys Pro Glu Pr Gly Pro Ala Ala Gly Ser 275 280 285

Gln Val Met Ala Trp Phe Arg Ala Lys Thr Ser Glu Ala Ser Asp Val 290 295 300

Met Ser Phe Leu Thr Cys Tyr His Phe Asp Leu His Trp Glu His His 305 310 315 320

Arg Trp Pro Phe Ala Pro Trp Trp Gln Leu Pro His Cys Arg Arg Leu 325 330 335

Ser Gly Arg Gly Leu Val Pro Ala Leu Ala 340 345

- 1. A DNA construct comprising in the 5' to 3' direction of transcription operably linked a promoter region directing transcription to the seed of an oilseed plant, a nucleotide sequence coding for at least one peptide with enzyme activity necessary for keto group containing xanthophyll production and esterification in an oilseed plant and a transcriptional termination region.
- 2. The DNA construct according to claim 1, which between the promoter region and the nucleotide sequence coding for at least one peptide with enzyme activity additionally comprises a nucleotide sequence coding for a transit peptide directing the translated fusion polypeptide to the chloroplast of the oilseed plant.
- 3. The DNA construct according to claim 1 or 2, wherein the promoter is a napin promoter, the peptide with enzyme activity necessary for keto group containing xanthophyll production and esterification is selected from the group consisting of peptides with, 1-Ddeoxyxylulose 5-phosphate synthase, isopentenyl pyrophosphate:dimethylallyl pyrophosphate isomerase, geranylgeranyl pyrophosphate synthase, phytoene synthase, phytoene desaturase, zeta-carotene desaturase, lycopene beta-cyclase, β-carotene hydroxylase, β-carotene C-4oxygenase, and acyl transferase activity.
- 4. The DNA construct according to any one of claims 1 3, wherein the nucleotide sequence coding for a peptide with enzyme activity is a nucleotide sequence coding for a N-terminally truncated \(\beta\)-carotene C-4-oxygenase gene from the alga Haematococcus pluvialis.
- 5. The DNA construct according to claim 4, wherein the nucleotide sequence is SEQ ID NO:1.
- 25 6. Transgenic oilseed plant cell comprising the DNA construct of any one of claims 1-5.
  - 7. Transgenic oilseed plant cell according to claim 6, wherein the oilseed plant is selected from the group consisting of rape, sunflower, soybean and mustard.
    - 8. Transgenic oilseed plant-produced xanthophyll.
    - 9. Transgenic oilseed plant-produced canthaxanthin
    - 10. Transgenic oilseed plant-produced astaxanthin.
    - 11. Transgenic oilseed plant-produced astaxanthin esters.

10

15

20



Napin promoter **AAGCTTTCTTCATCGGTGATTGATTCCTTTAAAGACTTATGTTTCTTATCTTGCTTCTGA** GGCAAGTATTCAGTTACCAGTTACCACTTATATTCTGGACTTTCTGACTGCATCCTCATT TTTCCAACATTTTAAATTTCACTATTGGCTGAATGCTTCTTCTTTGAGGAAGAAACAATT CAGATGGCAGAAATGTATCAACCAATGCATATATACAAATGTACCTCTTGTTCTCAAAAAC **ATCTATCGGATGGTTCCATTTGCTTTGTCATCCAATTAGTGACTACTTTATATTTCAC** TCCTCTTTATTACTATTTCATGCGAGGTTGCCATGTACATTATATTTGTAAGGATTGAC **GCTATTGAGCGTTTTTCTTCAATTTTCTTTATTTTAGACATGGGTATGAAATGTGTGTTA** GAGTTGGGTTGAATGAGATATACGTTCAAGTGAAGTGGCATACCGTTCTCGAGTAAGGAT GACCTACCCATTCTTGAGACAAATGTTACATTTTAGTATCAGAGTAAAATGTGTACCTAT **AACTCAAATTCGATTGACATGTATCCATTCAACATAAAATTAAACCAGCCTGCACCTGCA** TCCACATTTCAAGTATTTTCAAACCGTTCGGCTCCTATCCACCGGGTGTAACAAGACGGA TTCCGAATTTGGAAGATTTTGACTCAAATTCCCAATTTATATTGACCGTGACTAAATCAA CTTTAACTTCTATAATTCTGATTAAGCTCCCAATTTATATTCCCCAACGGCACTACCTCCA TATGAAGTTAAGTTTTACCTTGTTTTTAAAAAGAATCGTTCATAAGATGCCATGCCAGA ACATTAGCTACACGTTACACATAGCATGCAGCCGCGGAGAATTGTTTTTCTTCGCCACTT **GTGCATGCATTATTACACGTGATCGCCATGCAAATCTCCTTTATAGCCTATAAATTAACT** CATCCGCTTCACTCTTTACTCAAACCAAAACTCATCAATACAAACAAGATTAAAAAACATA

End -28 untranslated leader TP start

CACGAGGATCCTCAGTCACACAAAGAGTAAAGAAGAACAATGGCTTCCTCTATGCTCTCT

M A S S M L S

TCCGCTACTATGGTTGCCTCTCCGGCTCAGGCCACTATGGTCGCTCCTTTCAACGGACTT
S A T M V A S P A Q A T M V A P F N G L

AAGTCCTCCGCTGCCTTCCCAGCCACCCGCAAGGCTAACAACGACATTACTTCCATCACA
K S S A A F P A T R K A N N D I T S I T

TP End C-4-Oxygenase AGCAACGGCGGACGCGTTAACTGCATGTCTAGAATGCCATCCGAGTCGTCAGACGCAGCT SNGGRVNCMSRMPSESSDAA CGTCCTGCGCTAAAGCACGCCTACAAACCTCCAGCATCTGACGCCAAGGGCATCACGATG R P A L K H A Y K P P A S D A K G I T M GCGCTGACCATCATTGGCACCTGGACCGCAGTGTTTTTACACGCAATATTTCAAATCAGG ALTIGTWTAVFLHAIFQ'IR CTACCGACATCCATGGACCAGCTTCACTGGTTGCCTGTGTCCGAAGCCACAGCCCAGCTT L P T S M D Q L H W L P V S E A T A Q L TTGGGCGGAAGCAGCCTACTGCACATCGCTGCAGTCTTCATTGTACTTGAGTTCCTG LGGSSSLLHIAAVFIVLEFL TACACTGGTCTATTCATCACCACACATGACGCAATGCATGGCACCATAGCTTTGAGGCAC Y T G L F I T T H D A M H G T I A L R H AGGCAGCTCAATGATCTCCTTGGCAACATCTGCATATCACTGTACGCCTGGTTTGACTAC ROLNDLLGNICISLYAWFDY AGCATGCTGCATCGCAAGCACTGGGAGCACCACAACCATACTGGCGAAGTGGGGAAAGAC S M L H R K H W E H H N H T G E V G K D CCTGACTTCCACAAGGGAAATCCCGGCCTTGTCCCCTGGTTCGCCAGCTTCATGTCCAGC PDFHKGNPGLVPWFASFMSS TACATGTCCCTGTGGCAGTTTGCCCGGCTGGCATGGTGGGCAGTGGTGATGCAAATGCTG YMSLWQFARLAWWAVVMOML GGGGCGCCCATGGCAAATCTCCTAGTCTTCATGGCTGCAGCCCCAATCTTGTCAGCATTC G A P M A N L L V F M A A A P I L S A F CGCCTCTTCTACTTCGGCACTTACCTGCCACACAAGCCTGAGCCAGGCCCTGCAGCAGGC RLFYFGTYLPHKPEPGPAAG TCTCAGGTGATGGCCTGGTTCAGGGCCAAGACAAGTGAGGCATCTGATGTGATGAGTTTC SQVMAWFRAKTSEASDVMSF CTGACATGCTACCACTTTGACCTGCACTGGGAGCACCACAGATGGCCCTTTGCCCCCTGG LTCYHFDLHWEHHRWPFAPW C-4 oxygenase Stop WQLPHCRRLSGRGLVPALA\*

FIG.1 (cont.)

:::;



Fig.1 (cont.)

## 29295/BN

# **Abstract**

A DNA construct comprising in the 5' to 3' direction of transcription operably linked a promoter region directing transcription to the seed of an oilseed plant, a nucleotide sequence coding for at least one peptide with enzyme activity necessary for keto group containing xanthophyll production and esterification in an oilseed plant and a transcriptional termination region is disclosed. The DNA construct may additionally comprise a nucleotide sequence coding for a transit peptide directing the translated fusion polypeptide to the chloroplast of the oilseed plant. The peptide with enzyme activity is preferably a peptide with β-carotene C-4-oxygenase activity, e.g. from the alga *Haematococcus pluvialis*.

Comprised by the invention are also a transgenic oilseed plant cell, e.g. of rape, sunflower, soybean or mustard origin; transgenic oilseed plant-produced xanthophyll; transgenic oilseed plant-produced canthaxanthin; transgenic oilseed plant-produced astaxanthin; and transgenic oilseed plant-produced astaxanthin esters.

15

10